

ANTHELMINTIC POTENTIAL OF CRUDE ETHANOLIC EXTRACTS OF SELECTED PLANT SPECIES

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ABSTRACT

The present article illustrates the Anthelmintic activity of crude ethanolic extracts of Azadirachta indica, Phyllanthus emblica, Carica papaya and Citrus limetta leaves. The anthelmintic assay was carried out on Indian adult earthworms (Pheretima Posthuma), as per the method of Ajaiyeoba et al., (2001) with slight modifications. The minimum time for paralysis/death of the worms, was recorded in the ethanolic extract of Citrus limetta leaves, followed by ethanolic extracts of Azadirachta indica + Carica papaya leaves (2:3 dose proportion), at 50 ppm concentration. The experimental evidences have demonstrated significant anthelmintic activity of ethanolic extracts of Azadirachta indica, Phyllanthus emblica, Carica papaya and Citrus limetta leaves, at selected optimized concentration and dose proportion combinations. Thus, selected plant species could be explored further, in terms of biochemical characterization of active molecules, coupled with dose proportion studies, that can contribute to anthelmintics with enhanced efficacy.

KEYWORDS: Anthelmintic Activity, Ethanolic Extracts, Earthworms, Azadirachta Indica, Phyllanthus Emblica, Carica Papaya & Citrus Limetta

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INTRODUCTION

Helminthes infections, known as Helminthiasis is one of the most important animal diseases worldwide (Kosalge and Fursule, 2009; Roy *et al.*, 2012; Mehta *et al.*, 2012; Dogar *et al.*, 2012; Vennila *et al.*, 2015) and probably causes more morbidity, greater economic and social deprivation, among humans and animals, than any single group of parasites, thus distressing a large proportion of world's population (Partap *et al.*, 2012). Soil transmitted Helminthiasis (STHs) affects an average of 24% people, worldwide (WHO Fact Sheets, updated 2017). Helminthiasis inflicting heavy production losses in grazing/farm animals (Dogar *et al.*, 2012; Roy *et al.*, 2012; Goswami *et al.*, 2013). Helminthiasis is more widespread in developing countries, probably due to poor sanitation, malnutrition, and poor hygiene maintenance, etc. (Wath *et al.*, 2014). Helminthiasis treatment required ideal drugs, that should offer high efficacy, broad spectrum activity, free from toxic effects and cost effectiveness (Goswami *et al.*, 2013). Anthelmintics are drugs, that either kill or expel parasitic worms (Tripathi, 2004; Pillai and Nair, 2011; Mehta *et al.*, 2012). The problems associated with the use of chemical drugs like higher cost (Vennila *et al.*, 2015), several side effects (Gogoi *et al.*, 2014; Bochalaet *et al.*, 2016) and development of drug resistance (Mascie-Taylor and Karin, 2003; Wath *et al.*, 2014; Vennila *et al.*, 2015) could pave the way for herbal remedies, as alternative anthelmintics (Ashok Kumar *et al.*, 2010; Yadav and Singh, 2011; Dogar *et al.*, 2012; Goswami *et al.*, 2013; Wath *et al.*, 2014; Vennila *et al.*, 2015). Herbal medicines offer ease of use (Ullahet *et al.*, 2013), negligible side effects

(Mehta *et al.*, 2012; Ullah *et al.*, 2013), better potency (Mehta *et al.*, 2012) and cost effectiveness (Ullah *et al.*, 2013; Wath *et al.*, 2014), in recent years. The traditional medicines hold a great promise, as a source of effective anthelmintic, particularly in tropical developing countries, like India (Vennila *et al.*, 2015). Despite the prevalence of parasitic infections, the research on anthelmintic drug is poor (Aswaret *et al.*, 2008). Therefore, in the current study, evaluation of anthelmintic activity of crude ethanolic extracts of selected plant species (*Azadirachta indica*, *Phyllanthus emblica*, *Carica papaya* and *Citrus limetta* leaves) has been reported.

RESEARCH METHODOLOGY

For In-vitro studies, Earthworms proved to be good test worms, because of their longer survival in PBS (Dogar *et al.*, 2012), easy accessibility (Vennila *et al.*, 2015), and due to its anatomical and physiological resemblance with the intestinal roundworm parasites of human beings (Vidyarthi, 1977; Thorn *et al.*, 1977; Vigar, 1984). A/c to Goswami *et al.*, (2013), among worms, 18% of Earthworm i.e. *Pheretima Posthuma*, have been reported as targeted parasites for the evaluation of anthelmintic activity. Therefore, it was decided to carry out the In-vitro anthelmintic activity, on Indian adult earthworms (*Pheretima Posthuma*) as preliminary assay. The healthy adult earthworms were collected from the local nursery. The earthworms were maintained under normal Vermicomposting medium, with adequate supply of nourishment and water (Nisha *et al.*, 2012). The earthworms were firstly washed with normal saline, to remove all faecal matter (Mute *et al.*, 2009; Ashok Kumar *et al.*, 2010). Earthworms 6-8 cm in length (Kosalge and Fursule, 2009) and 0.2-0.3 cm in width (Nisha *et al.*, 2012; Mute *et al.*, 2009), 0.25-0.35 g in weight were used, for all experimental protocols.

Plant materials (Fresh leaves of *Azadirachta indica*, *Phyllanthus emblica*, *Carica papaya* and *Citrus limetta*) were cleansed with distilled water and allowed to dry for 15 days, under darkness. Followed to this, it was coarsely crushed using homogenizer and grinded mechanically of mesh size 1 mm. The powdered plant material was extracted with 70% Ethanol. After 48 hours, extracts were filtered by using muslin cloth (cheese cloth) followed by Whatman filter paper No.1 and filtrates were evaporated till dryness and weighed. The crude extracts were stored in air tight glass containers at 4°C till further analysis (Bibi *et al.*, 2011; Shobowale *et al.*, 2013).

Percent of the yield of Extract was calculated according to Patil and Gaikwad (2010); Ekaluo *et al.*, (2015). The anthelmintic assay was carried out as per the method of Ajaiyeoba *et al.*, (2001) with slight modifications. Solvents used in extraction and Reagents for phytochemical and anthelmintic analysis was of pure analytical grade. All the results were expressed as a mean \pm SEM of six worms in each group.

RESULTS AND DISCUSSIONS

Percentage Yield of Extract

Percent yield of the crude ethanolic extracts from *Azadirachta indica*, *Phyllanthus emblica*, *Carica papaya* and *Citrus limetta* leaves after complete extraction were 11.91 %, 4.33 %, 8.81 % and 8.41 %, respectively. Highest yield was observed in the ethanolic extract of *Azadirachta indica* leaves followed by ethanolic extracts of *Carica papaya* and *Citrus limetta* leaves. Similar results were also reported in the previous studies. Kazeem *et al.*, (2013) found, extract yield of 4.35 % and 2.25 % in aqueous and ethanolic extracts of *Azadirachta indica* leaves, respectively. A/c to Sithisarn *et al.*, (2006), Maceration, Percolation and Soxhlet extraction of *Azadirachta indica* leaves with concentrations of ethanol ranges from 20-95%, gave a crude extract yield b/w 10-25% w/w. Chavez-Quintal *et al.*, (2011) in his studies, observed the yield of ethanolic extracts of *Carica papaya* leaves in the range of 3.1-5.9 % under variable extraction time and flour: solvent ratio.

Gitika and Kumar (2016) were reported % extract yield of 10.52 and 15.41 in ethanollic and aqueous extracts of *Phyllanthus emblica* leaves, respectively. 19.25 % of extract yield was observed by Muthiah *et al.*, (2012) in 60% ethanollic extract of *Citrus limetta* leaves and peels.

Anthelmintic Activity

In order to investigate the anthelmintic properties of *Azadirachta indica*, *Phyllanthus emblica*, *Carica papaya* and *Citrus limetta* leaves, the present study has been conducted with ethanollic extracts optimized at different concentrations and dose proportion combinations. Albendazole (at 10, 15 & 20 mg/ml) served as standard throughout the experiment. Ethanollic extracts of *Azadirachta indica*, *Phyllanthus emblica*, *Carica papaya* and *Citrus limetta* leaves produced a significant anthelmintic activity in a dose dependent manner. It was observed that the ethanollic extract of *Carica papaya* leaves has a more profound effect as anthelmintic, than the ethanollic extract of *Azadirachta indica* leaves at 50 ppm concentration. The higher anthelmintic potential of mixed dilutions of *Azadirachta indica* and *Carica papaya* at the dose proportion of 2:3 over 3:2 dose proportion at 50 ppm concentration also justified the same. However, minimum time for paralysis/death of the worms were recorded in the ethanollic extract of *Citrus limetta* leaves, as well as in mixed dilutions of *Azadirachta indica* and *Carica papaya* (2:3 dose proportion), at 50 ppm concentration.

In earlier studies, Aggarwal and Bagai (2014), confers that, ethanollic and aqueous extracts of *A. indica* leaf may act as potential Vermifuge or Vermicide. Roy *et al.*, (2012) reported the significant anthelmintic property of hydro-alcoholic and chloroform extracts of the stem and leaves of *Carica papaya* at 5%, 2.5% and 1% extract concentrations, against *Pheretima Posthuma*. Sukanya *et. al.*, (2013) studied anthelmintic activity of methanol, butanol, ethyl acetate and aqueous extracts of *Phyllanthus emblica* leaves & bark, against *Pheretima Posthuma*. Barrage *et al.*, (2011), demonstrated anthelmintic activity of petroleum ether extract of *Citrus medica* (L.) leaves, against *Pheretima Posthuma*.

CONCLUSIONS

The experimental evidences have demonstrated significant anthelmintic activity of ethanollic extracts of *Azadirachta indica*, *Phyllanthus emblica*, *Carica papaya* and *Citrus limetta* leaves, as they took less time to cause paralysis and death of the earthworms, at selected optimized concentration and dose proportion combinations. The present findings were justified from the comparative analysis with standard drug Albendazole, at different doses. However, the findings of present study did not exclude the possibility that, the ethanollic extracts of studied plant species under variable concentrations and dose proportion combinations, having lower anthelmintic activity, might be efficacious against other species of helminths. It is recommended that, the selected plant species could be explored further, in terms of biochemical characterization of active molecules, coupled with dose proportion studies that can contribute to anthelmintics with enhanced efficacy.

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REFERENCES

1. Aggarwal, R. and Bagai, U. (2014). Evaluation of anthelmintic activity of ethanolic and aqueous leaf extracts of *Azadirachta indica* on phosphatases in *Gastrothylax indicus*. *IOSR Journal of Pharmacy and Biological Sciences*, 9(5): 98-104.
2. Ajaiyeoba, E.O., Onocha, P.A. and Olarenwaju, O.T. (2001). In-vitro anthelmintic properties of *Buchholziacoriaceae* and *Gynandropsis gynandra* extract. *Pharm Biol.*, 39: 217.
3. Ashok Kumar, B.S., Lakshman, K., Jayaveera, K.N., Nandeesh, R., Manoj, B. and Ranganayakulu, D. (2010). Comparative In-vitro anthelmintic activity of three plants from the *Amaranthaceae* family. *Arch. Biol. Sci., Belgrade.*, 62(1): 185-189.
4. Aswar, M., Aswar, U., Watkar, B., Vyas, M., Wagh, A. and Gujar, K.N. (2008). Anthelmintic activity of *Ficus benghalensis*. *International Journal of Green Pharmacy*, 170-172.
5. Bairagi, G.B., Kabra, A.O. and Mandade, R.J. (2011). Anthelmintic Activity of *Citrus medica* (L.) leaves in Indian Adult Earthworm. *International Journal of PharmTech Research*, 3(2): 664-667.
6. Bibi, Y., Nisa, S., Chaudhary, F. M. and Zia, M. (2011). Antibacterial activity of some selected medicinal plants of Pakistan. *BMC Complementary and Alternative Medicine*, 11: 52. <http://doi.org/10.1186/1472-6882-11-52>.
7. Bochala, V.J., Simham, S., Katakam, S., Punuru, P. and Shaik, C.B. (2016). Anthelmintic activity of methanolic extract of *Peltophorum pterocarpum* leaves. *World Journal of Pharmacy and Pharmaceutical sciences*, 5(2): 1036-1042.
8. Chavez-Quintal, P., Gonzalez-Flores, T., Rodriguez-Buenfil, I. and Gallegos-Tintore, S. (2011). Antifungal Activity in Ethanolic Extracts of *Carica papaya* L. cv. Maradol Leaves and Seeds. *Indian J Microbiol.*, 51(1): 54-60.
9. Neetu Singh et al., Phytochemical Analysis of Plant Species of Genus *Zanthoxylum*, *International Journal of Medicine and Pharmaceutical Sciences (IJMPS)*, Volume 6, Issue 1, January - February 2016, pp. 1-8
10. Dogar, Z.H.M., Faiza, A., Khalil-ur-Rahman, Shazia, T. and Abdul, R., et al. (2012). In vivo and in vitro antihelmintic activity of gemmotherapeutically treated *Azadirachta indica* (Neem) against gastrointestinal nematodes of sheep and earthworms. *African J Pharm Pharmacol.*, 6(46): 3171-3179.
11. Ekaluo, U.B., Ikpeme, E.V., Udensi, O.U., Ekerette, E.E., Usen, S.O. and Usoroh, S.F. (2015). Comparative in vitro Assessment of Drumstick (*Moringa oleifera*) and Neem (*Azadirachta indica*) Leaf Extracts for Antioxidant and Free Radical Scavenging Activities. *Research Journal of Medicinal Plant*, 9(1): 24-33.
12. Gitika and Kumar, M. (2016). Evaluation of antibacterial activity of *Phyllanthus emblica* (L.) leaves extracts against gram-positive and gram-negative bacteria. *World Journal of Pharmaceutical Research*, 5(8): 1459-1470.
13. Gogoi, B., Kakoti, B.B., Bora, N.S. and Yadav, P. (2014). In vitro antihelmintic activity of bark extract of *Cinnamomum bejolghota* (Buch-Ham.) in Indian adult earthworm (*Pheretima posthuma*). *Asian Pac J Trop Dis.*, 4(Suppl 2): S924-S927.
14. Goswami, S., Nishad, S., Rai, M., Madhesiya, S., Malviya, A., Pandey, P., Gautam, V. and Yadav, S. (2013). Plant seeds used for Anthelmintic activity: A Review. *Indian Journal of Research in Pharmacy and Biotechnology*, 1(4): 533-536.
15. Kazeem, M.I., Dansu, T.V. and Adeola, S.A. (2013). Inhibitory effect of *Azadirachta indica* A. Juss Leaf extract on the activities of α -amylase and α -glucosidase. *Pakistan Journal of Biological Sciences*, pp:1-5.
16. Kosalge, S.B. and Fursule, R.A. (2009). Investigation of In-vitro Anthelmintic activity of *Thespesia lampas* (CAV.). *Asian Journal of Pharmaceutical and Clinical Research*, 2(2): 69-71.
17. Mascie-Taylor, C.G.N. and Karin, E. (2003). The burden of chronic disease. *Science*, 302: 1921-1922.
18. Mehta, P., Phutane, S. and Sutar, S. (2012). In-vitro anthelmintic activity of whole plant of *Ventilago denticulata* willd. against

- Pheretima posthuma*. Asian Journal of Pharmaceutical and Clinical Research, 5(3): 200-201.
19. Mute, V.M., Sampat, V.M., Patel, K.A., Sanghavi, K., Mirchandani, D. and Babaria, P.C. (2009). Anthelmintic effect of *Tamariznd indica* (Linn.) leaves juice extract on *Pheretima posthuma*. International Journal of Pharma Research and Development-Online, 7: 1-6. Publication Ref No.: IJPRD/2009/PUB/ARTI/VOL-7/SEP/001.
 20. Muthiah, P.L., Umamaheswari, M. and Asokkumar, K. (2012). In-vitro antioxidant activities of leaves, fruits and peel extracts of citrus. International Journal of Phytopharmacy, 2(1): 13-20.
 21. Nisha, P.V., Shruti, N., Sweta Swamy, K., Kumari, M., Vedamurthy, A.B., Krishna, V. and Hoskeri, J.H. (2012). Anthelmintic activity of *Pyrostegia venusta* using *Pheretima posthuma*. International Journal of Pharmaceutical Sciences and Drug Research, 4(3): 205-208.
 22. Partap, S., Kumar, S., Kumar, A., Sharma, N.K. and Jha, K.K. (2012). In-Vitro Anthelmintic Activity of *Luffa cylindrica* Leaves in Indian Adult Earthworm. Journal of Pharmacognosy and Phytochemistry, 1(2): 27-30.
 23. Patil, U.H. and Gaikwad, D.K. (2010). Phytochemical evaluation and bactericidal potential of *Terminalia arjunastem* bark. Int J Pharm Sci Res., 2(3): 614-619.
 24. Pillai, L. S. and Nair, B. R. (2011). A Comparative Study of the Anthelmintic Potential of *Cleome Viscosa* L. and *Cleome Burmanni* W. and A. Indian Journal of Pharmaceutical Sciences, 73(1): 98-100. <http://doi.org/10.4103/0250-474X.89766>.
 25. Roy, S.D., Goswami, R., Das, S., Shil, D., Baniya, R. and Haldar, S. (2012). Pharmacognostic Evaluation and Anthelmintic activity of Leaf and Stem Extract of *Carica papaya*. Journal of Pharmacy Research, 5(9): 4763-4766.
 26. Shobowale, O.O., Ogbulie, N.J., Itoandon, E.E., Oresgun, M.O., Olatope, S.O.A. (2013). Phytochemical and Antimicrobial Evaluation of Aqueous and Organic Extracts of *Calotropis procera* Ait Leaf and Latex. Nigerian Food Journal, 31(1): 77-82.
 27. Sithisarn, P., Supabphol, R. and Gritsanapan, G. (2006). Comparison of Free Radical Scavenging Activity of Siamese Neem Tree (*Azadirachta indica* A. Juss var. *siamensis* Valetton) Leaf Extracts Prepared by Different Methods of Extraction. Med Princ Pract., 15: 219-222.
 28. Sukanya M.K, Shimi, S. and Aruna, S.R. (2013). Phytochemical analysis, Antimicrobial screening and Antihelminthic properties of *Phyllanthus emblica*. Int J Pharm Bio Sci., 4(4): 55-64.
 29. Thorn, G.W., Admas, R.D., Brundwald, E., Isselbacher, K.J. and Petersdorf, R.G. (1977). *Harrisons Principles of Internal Medicine*. New York: McGraw Hill Co.
 30. Tripathi, K.D. (2004). *Essential of Medical pharmacology* 5th ed. Jaypee Brothers, pp: 759.
 31. Ullah, M., Khan, M.U., Mahmood, A., Malik, R.N., Hussain, M., Wazir, S.M., Daud, M. and Shinwari Z.K. (2013). An ethnobotanical survey of indigenous medicinal plants in Wana district south Waziristan agency, Pakistan. J Ethnopharmacol., 150(3): 918-24.
 32. Vennila, V., Selvarani, R., Anitha, R., Marthal, M.A., Nivetha, R. and Pavithra, V. (2015). In-vitro anthelmintic activity of extracts of *Erythrina variegata* leaves in Indian earthworm *Pheretima posthuma*. World Journal of Pharmacy and Pharmaceutical Sciences, 4(5): 1306-1318.
 33. Vidyarthi, R.D. (1977). *A Text book of Zoology*. 14th ed. New Delhi: S Chand and Co.
 34. Vigar, Z. (1984). *Atlas of Medical Parasitology*. 2nd ed. Singapore: P.G. Publishing House.
 35. Wath, M., Lakade, P. and Lande, P. (2014). Comparative evaluation of anthelmintic activity of two plants from the family *Euphorbiaceae*. Biolife, 2(2): 534-537.

36. WHO Fact Sheets. Soil-transmitted helminth infections. Updated on January 2017. Source: <http://www.who.int/mediacentre/factsheets/fs366/en/>.
37. Yadav, P. and Singh, R. (2011). A Review on anthelmintic drugs and their future scope. *Int J Pharm Pharm Sci*, 3(3): 17-21.